

# Mechanistic Modeling of Microbial Interactions: A Simplicity-Realism Trade-off

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**COMBINE 2018, Boston**

*October 8, 2018*

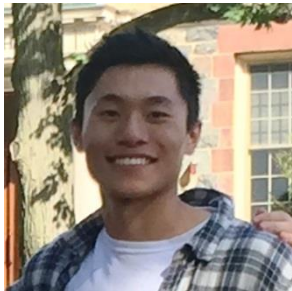


# Acknowledgment

## Experiments



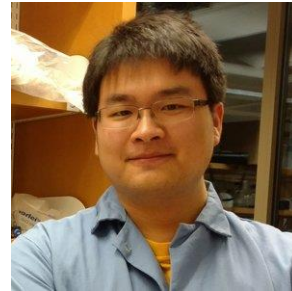
**Sam Dyckman**



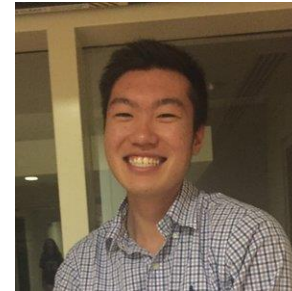
**Ray Lang**



**Catherine Henckel**



**Kevin Chen**



**David Fu**



**Sandra Dedrick**

## Simulations



**Suyen Espinoza**



**Ian Boland**



**Lori Niehaus**



**Minghao Liu**



**Kaitlin Chaung**

**Wenying Shou and Li Xie** (Fred Hutchinson Cancer Research Center)

**Michelle Meyer and Tim van Opijnen** (Boston College)

**Michael Shiaris** (UMass Boston)

**Shin Haruta** (Tokyo Metropolitan University)

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# Why are we interested in microbial communities?

**Microbial communities:** assemblies of interacting microbes

Why important? Impact on health, industry, and environment

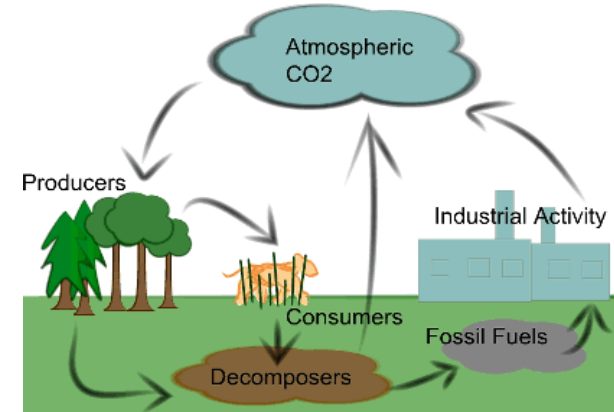
**Chronic wound infections**



**Waste treatment**



**Ecosystem carbon cycling**



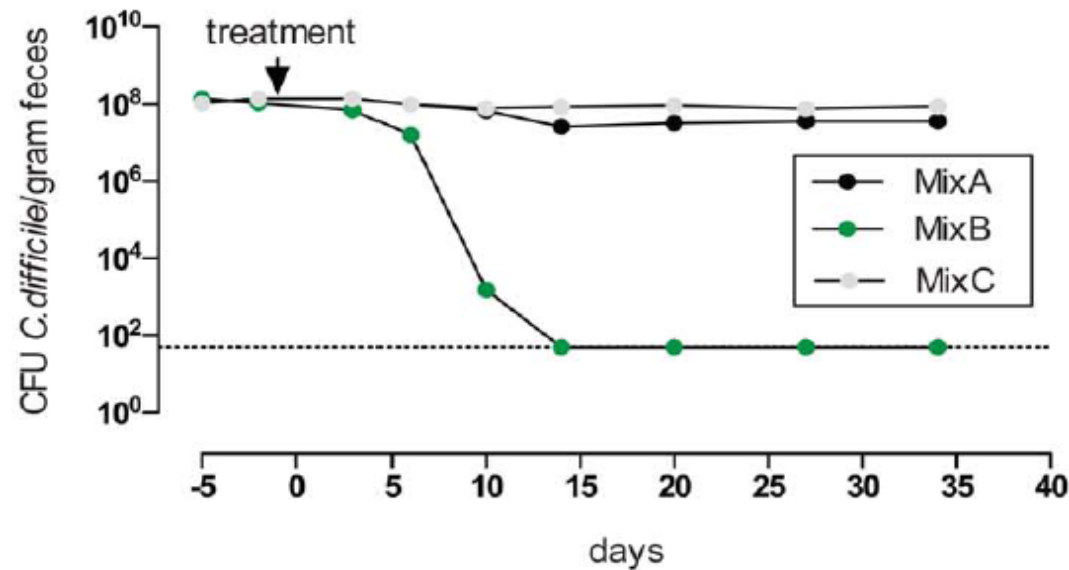
Long-term objective: control

- Eradicating harmful communities
- Maintaining useful communities

# Communities can exhibit functions not achievable by any of the individual species

## Example: Gut community can resist pathogens

- Constructed six-species community treats *C. diff* infection in mice
- No treatment with any of the single species (or other subsets)



Lawley et al, *PLOS Pathogens* (2012)

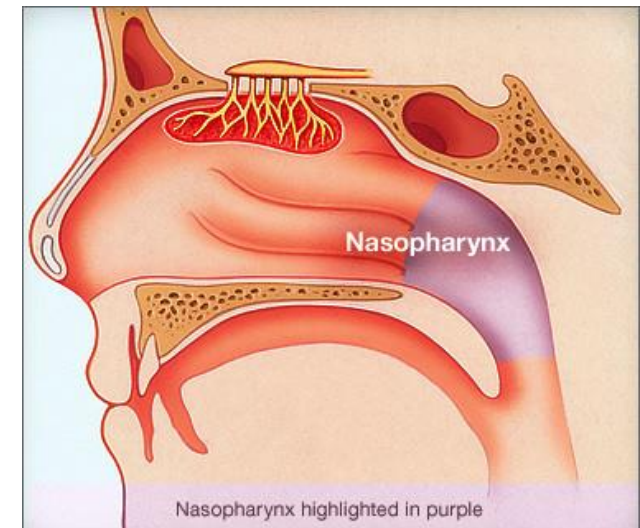
# Controlling communities requires a better understanding of underlying processes

## Example: nasal microbiota can prevent pathobiont colonization

- *Staph. aureus* present in ~30% of population (often not harmful)
- *S. aureus* carrier vs non-carrier states not dependent on host genes
- Possibly other harmless microbiota residents determine colonization
- Self-infection is the primary cause in hospitalized patients
- Use of antibiotics is not an effective solution

## Goal (to decrease the chance of infection):

- What are the processes that shape nasal microbiota?
- What strategy can convert carrier-type to non-carrier-type?





# Objective

**Understanding the basic processes that  
shape microbial communities (using simple models)**

with the goal of

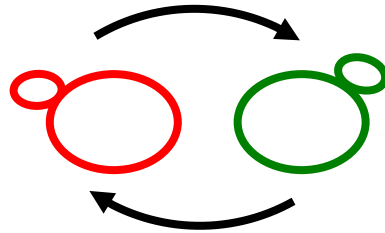
**devising strategies to control communities**

# Approaches for studying microbial communities

Natural

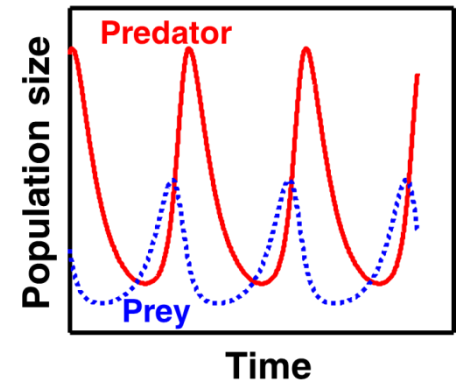


Engineered



Transparency  
Control

Simulated

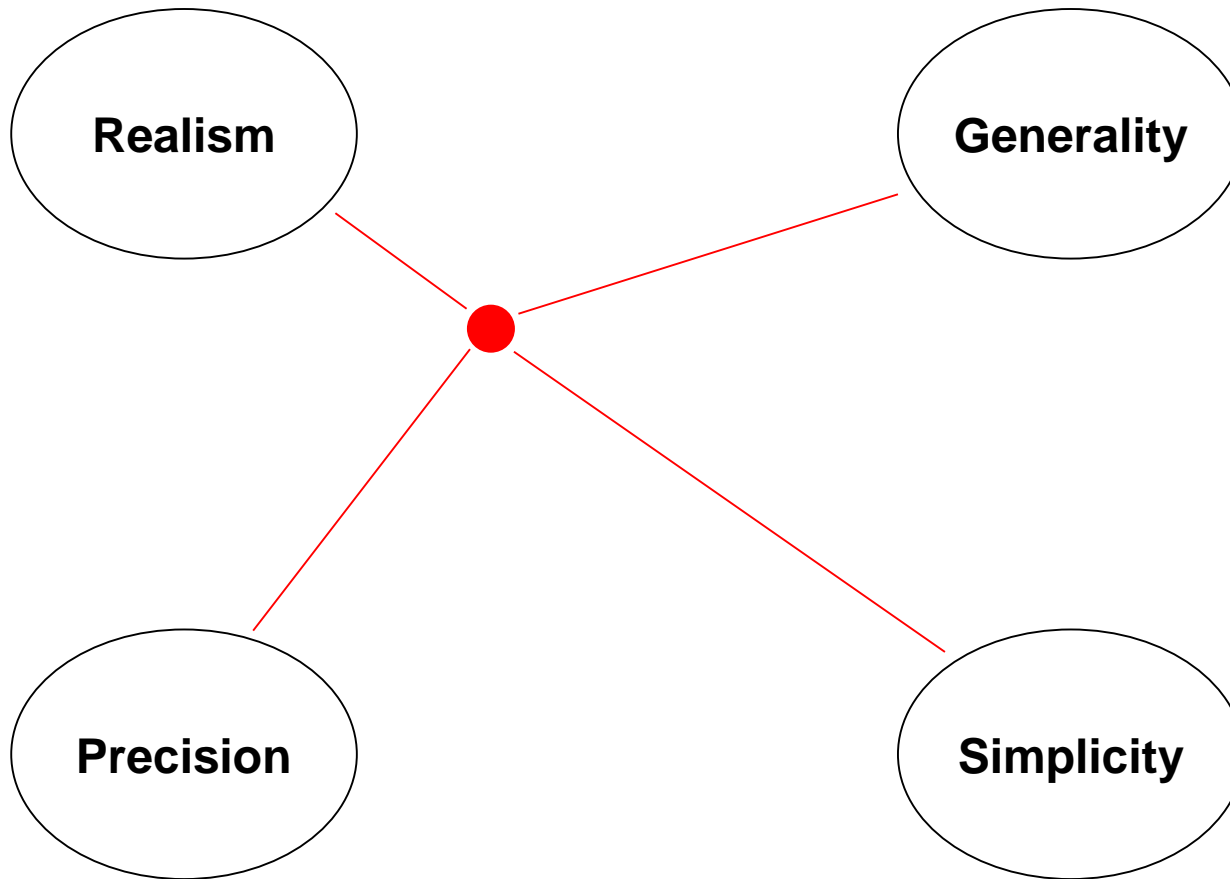


Realism  
Complexity

Informing models to make them more realistic,  
without sacrificing transparency/control

# Trade-off in building a model

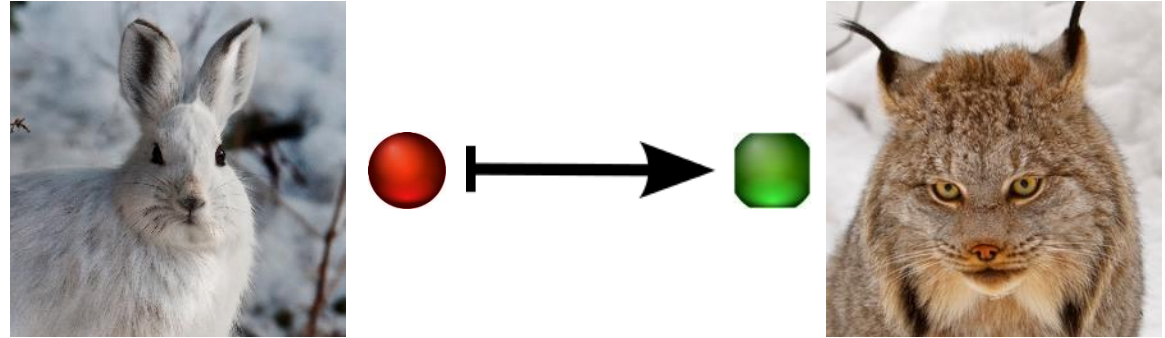
Modified from Levins 1966, *Strategy of model building in population biology*





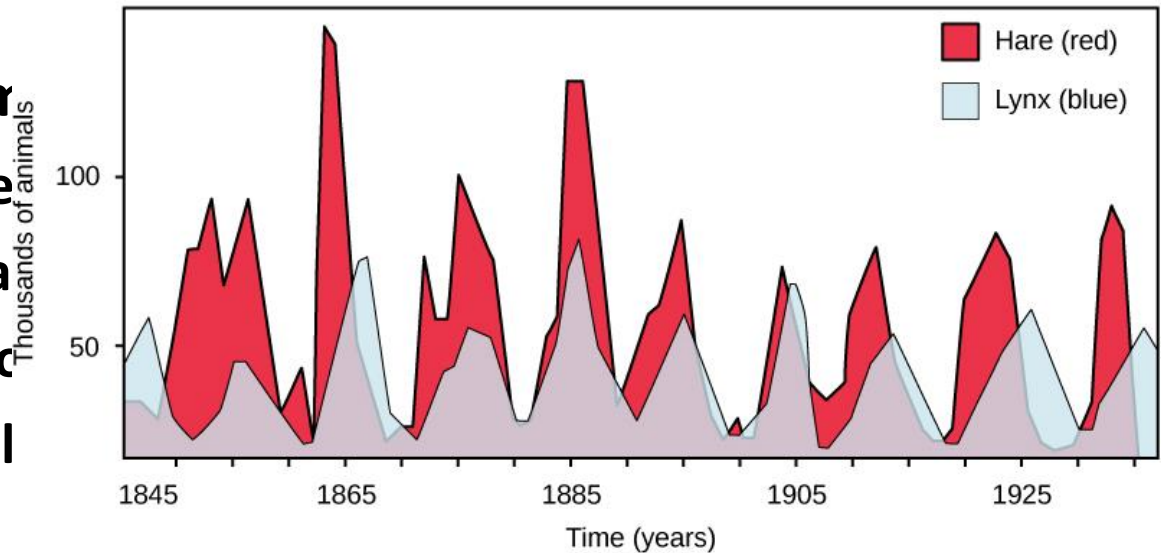
# Pairwise fitness models are most commonly used

Modeling interactions as net fitness effects, regardless of mechanisms  
e.g. Lotka-Volterra



## Advantages of pairwise models

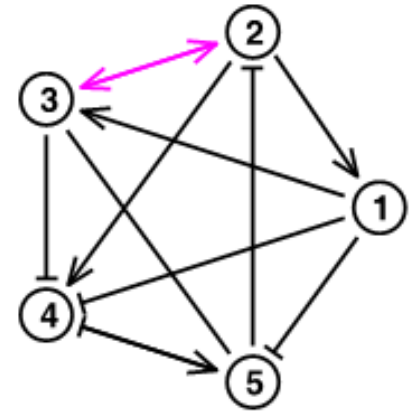
- No need to know interactions
- Easy to estimate parameters
- Some empirical support
- Easy to extend to multiple species



Is pairwise modeling applicable to microbial communities?

# Lotka-Volterra modeling of microbial communities

- Intrinsic assumptions
    - Pairwise interactions can be properly modeled
    - Interactions are independent
  - Many interactions among species are mediated through chemicals
    - Beneficial metabolic exchanges
    - Inhibiting metabolic byproducts, toxins, and antibiotics
- How do LV models handle interactions mediated through chemicals?



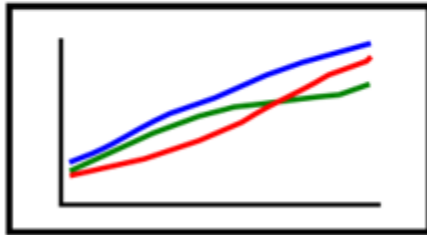
# Comparing predictions: pairwise versus mechanistic

Derive equivalent  
pairwise model

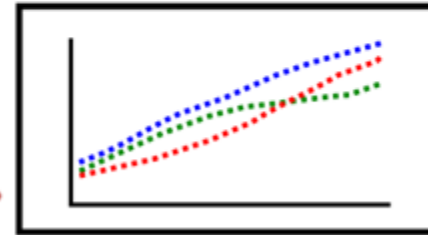
Mechanistic model  
“ground truth”

Pairwise model  
“common approximation”

Simulate dynamics



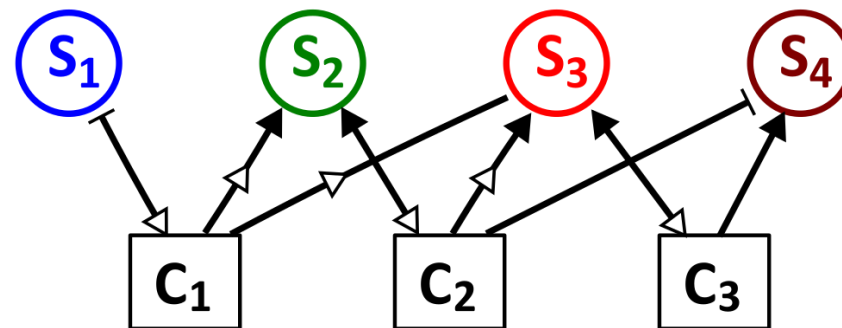
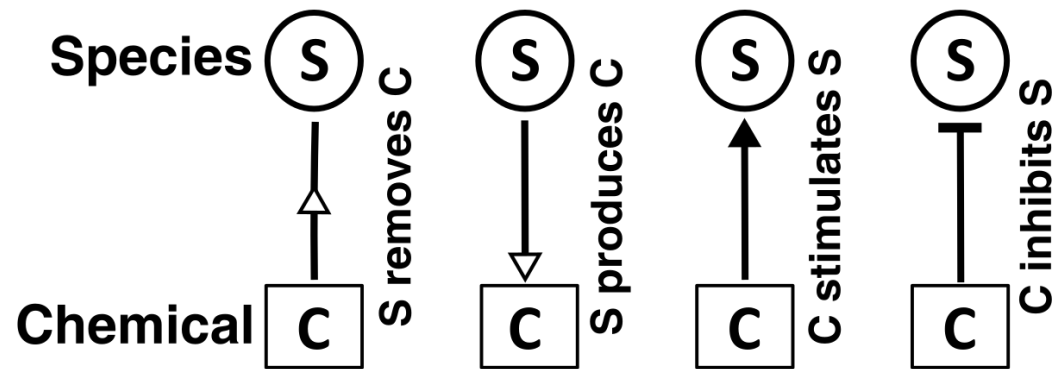
Simulate dynamics



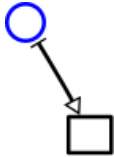
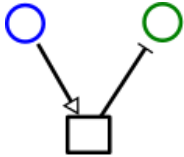
Compare predictions

# Mediator-explicit (ME) modeling

Explicitly incorporating chemical mediators of interactions in the model



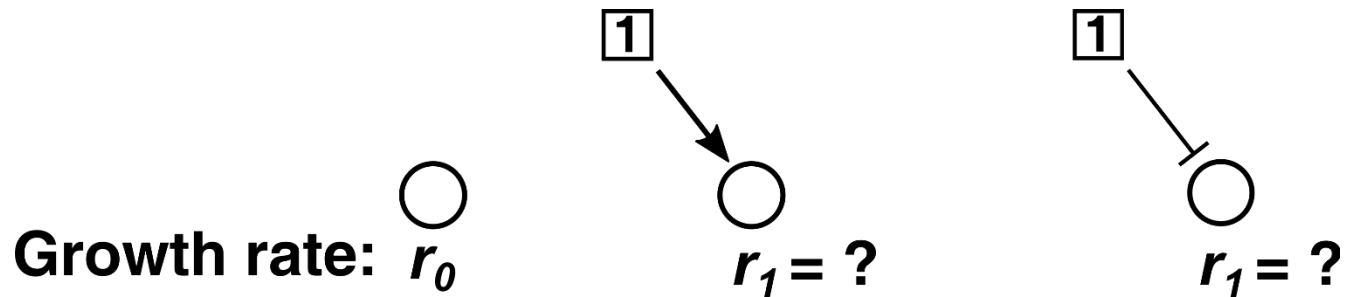
# Examples of interactions among microbial species

Interaction		Type	Example
Intrapopulation	Interpopulation		
		Target cell not degrading its inhibitor	Accumulation of end-products such as acetate (Kato et al. 2005) or ethanol (Gause 1934a)

# Assumptions of the mediator-explicit model

## How to model the effect of individual mediators?

- Measure cell's response to sample mediators



- Our “guinea pig”: *E. coli* MG1655 or environmental isolates *Brevibacillus agri* and *Pseudoxanthomonas taiwanensis*
- Choice of mediators:
  - Facilitators: carbon sources (e.g. glycerol); amino acids
  - Inhibitors: antibiotics (e.g. gentamicin); fermentation products (e.g. acetic acid)
- Read-out: growth rate in exponential phase



# Modeling the effect of growth facilitators

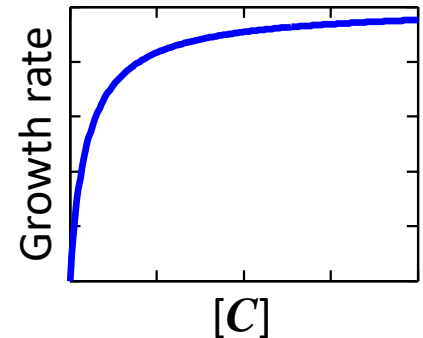
Assumptions:

- Single limiting resource,  $C$
- Michaelis-Menten uptake of resources
- Cells divide after acquiring enough of the limited resource

$$\frac{\Delta S}{\Delta t} = \frac{(\text{uptake rate per cell}) \times (\text{cell \#})}{\text{amount required to build a cell}} = \frac{v_M C / (C + K)}{\alpha} S$$

Or

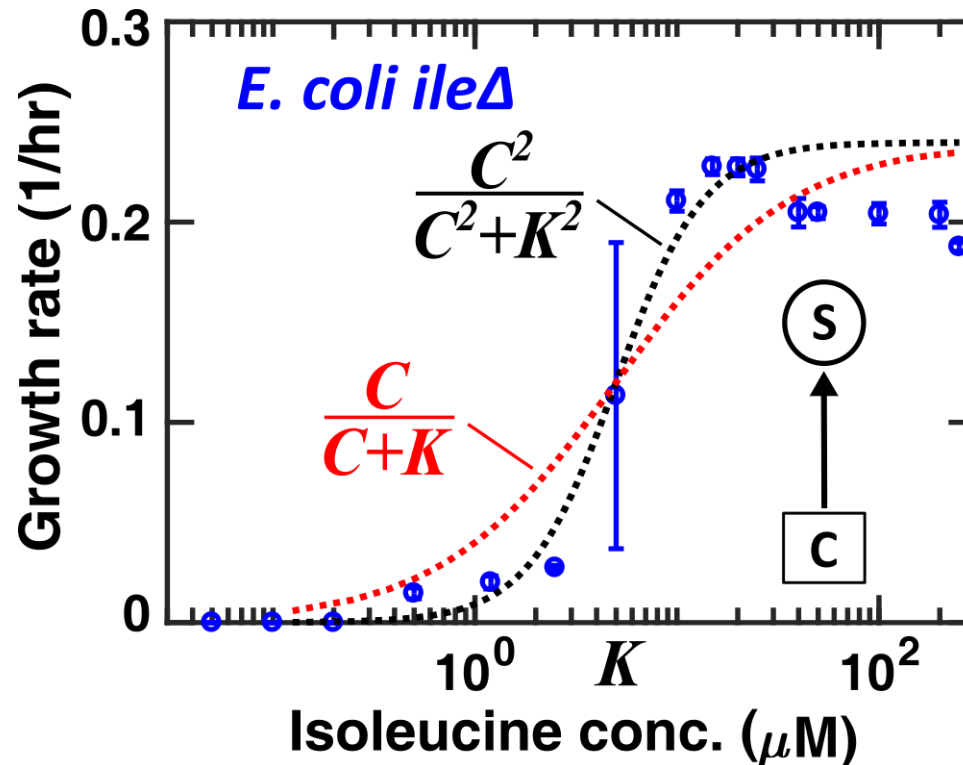
$$\frac{dS}{dt} = r_M \frac{C}{C + K} S$$



# Experimental test of the effect of single facilitators

Isoleucine facilitation (for isoleucine auxotrophic *E. coli*)

- Measured exponential growth rate at different concentrations



- Data suggests transition is often steeper:

$$\frac{dS}{dt} = r_M \frac{C^n}{C^n + K^n} S ; n > 1$$

# Modeling the effect of growth inhibitors

## Assumptions:

- Cell-inhibitor random encounter in a well-mixed environment
- Upon encounter, cells die with a fixed probability,  $p_d$
- Motivated by ecological models of prey-predators, chance of encounter per unit time is proportional to  $SC$  ( $S$ : cell density,  $C$ : inhibitor conc.)

Change in population size in unit time

= cells born – cells dead due to inhibitor encounter

$$\frac{\Delta S}{\Delta t} = r_0 S - p_d r_e SC$$

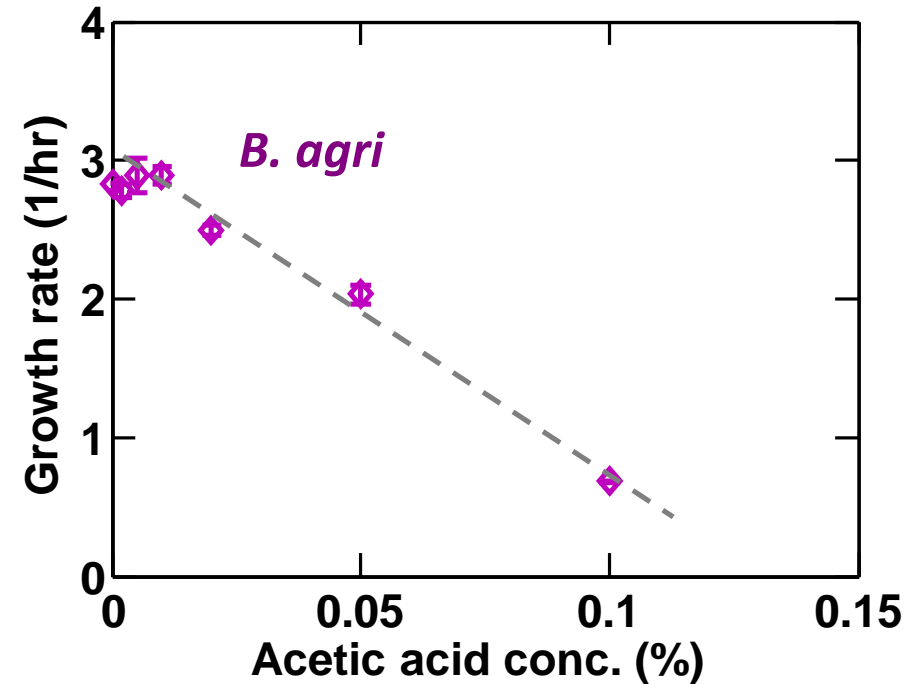
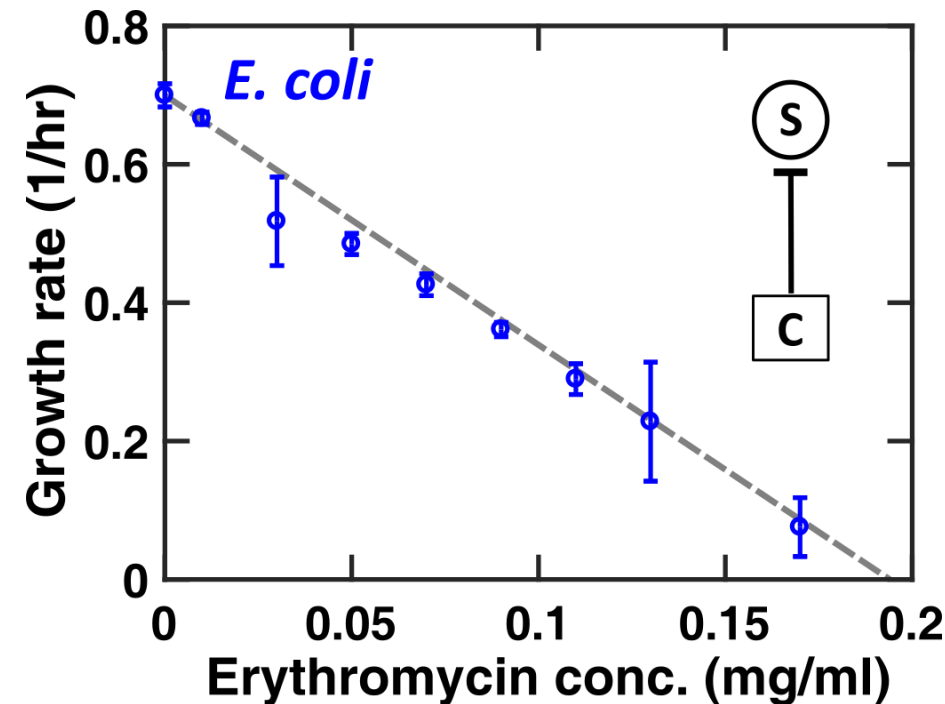
Or

$$\frac{dS}{dt} = (r_0 - \eta C) S$$

# Experimental test of the effect of single inhibitors

## Acetic acid and erythromycin inhibition

- Measured exponential growth rate at different concentrations

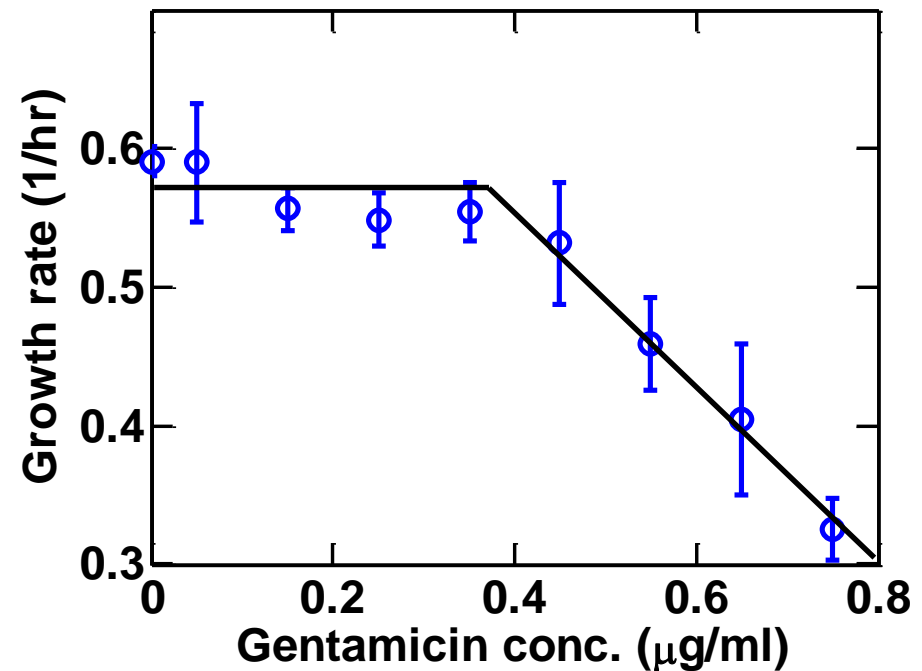
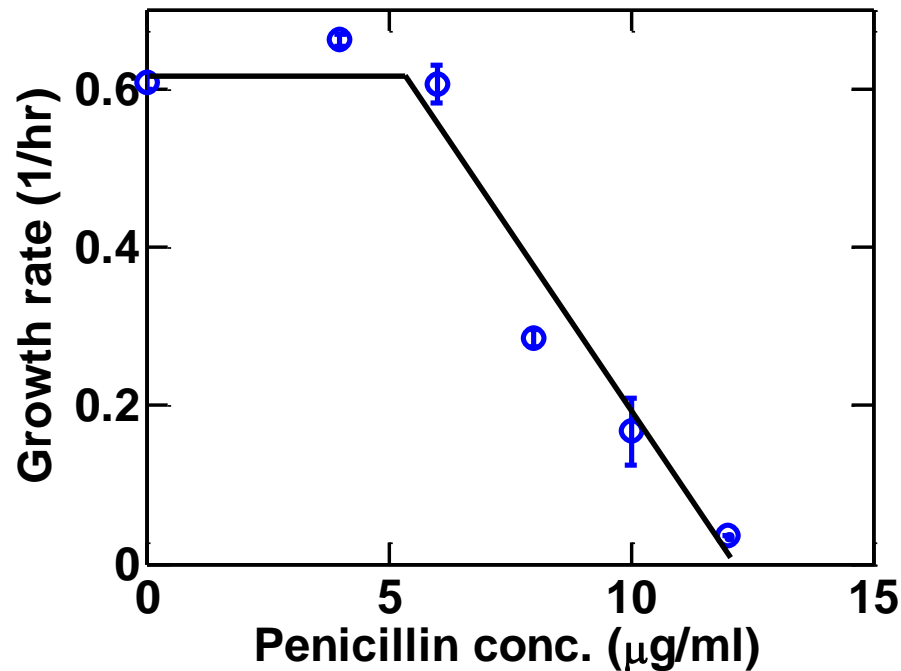


- In many cases:

$$\frac{dS}{dt} = (r_0 - \eta C) S$$

# Experimental test of the effect of single inhibitors

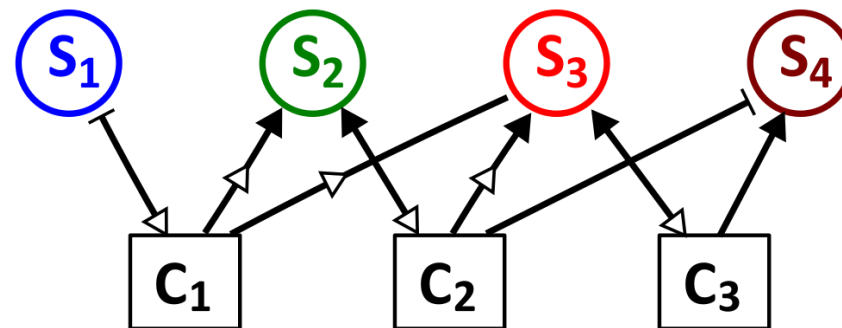
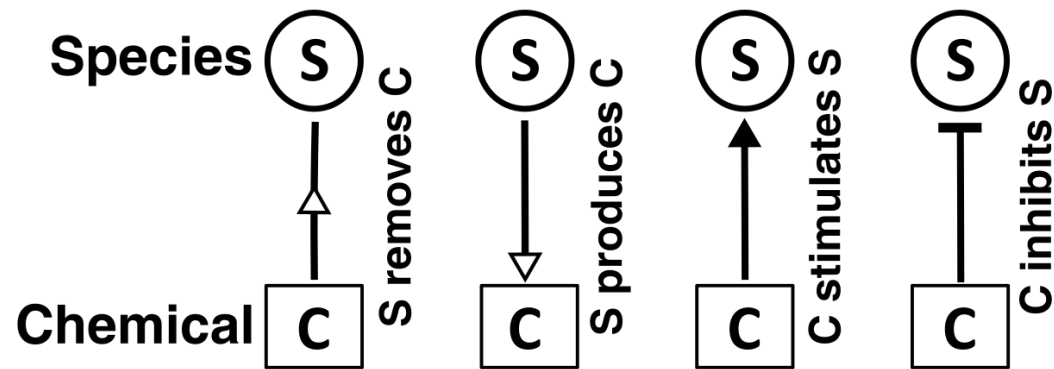
With some antibiotics, inhibition is effective beyond a threshold conc.



$$\frac{dS}{dt} = \begin{cases} r_0 S & ; C \leq C_{th} \\ r_0 S - \eta S (C - C_{th}) & ; C > C_{th} \end{cases}$$

# Mediator-explicit (ME) modeling

Explicitly incorporating chemical mediators of interactions in the model

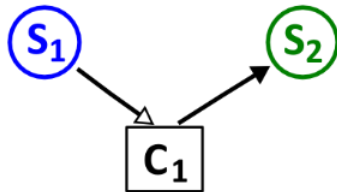




# LV does not capture chemical mediated interactions

Can the canonical LV model represent all microbial interactions? **X**

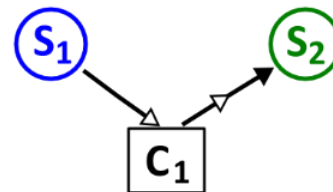
Reusable



$$\frac{dS_2}{dt} = r_{20} S_2 + r_{21} \frac{S_1}{S_1 + K_{21}} S_2$$

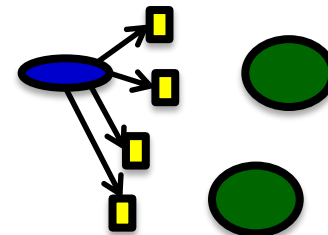
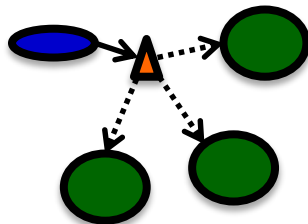
Saturable LV model

Consumable



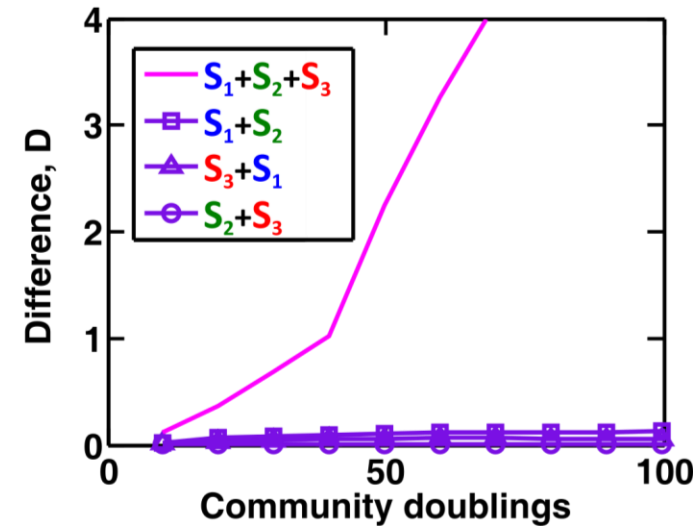
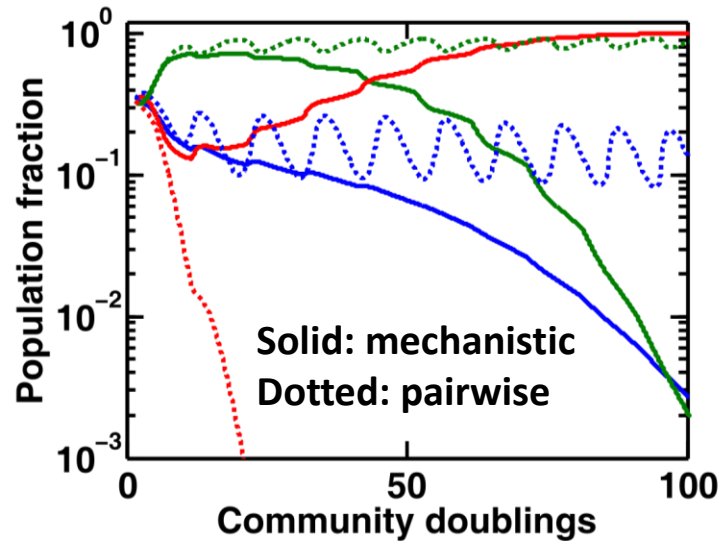
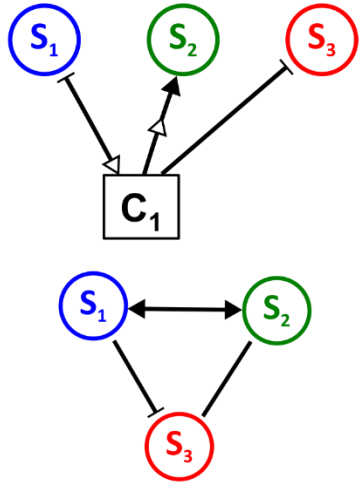
$$\frac{dS_2}{dt} \approx r_{20} S_2 + r_{S_2 C_1} \frac{S_1}{\omega S_1 + \psi S_2} S_2$$

Divided influence model



# LV does not capture chemical mediated interactions

Can the model be extended beyond two-species communities? **X**

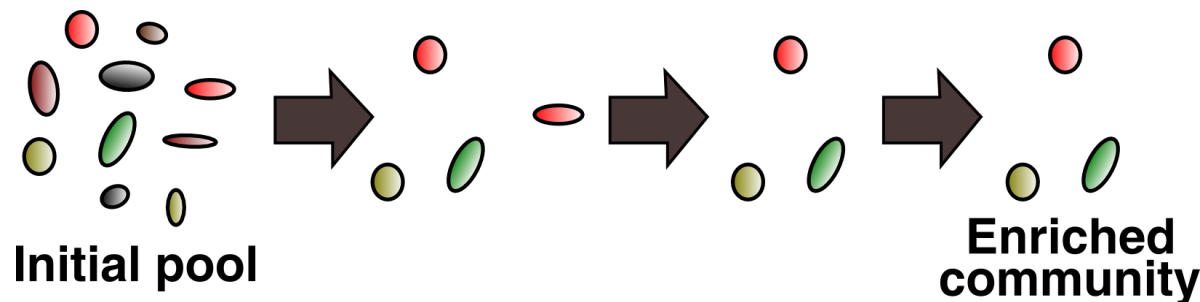


# Coexistence of species in microbial communities

How does the interaction network among species lead to their coexistence?

Insights from common features of simulated coexistence

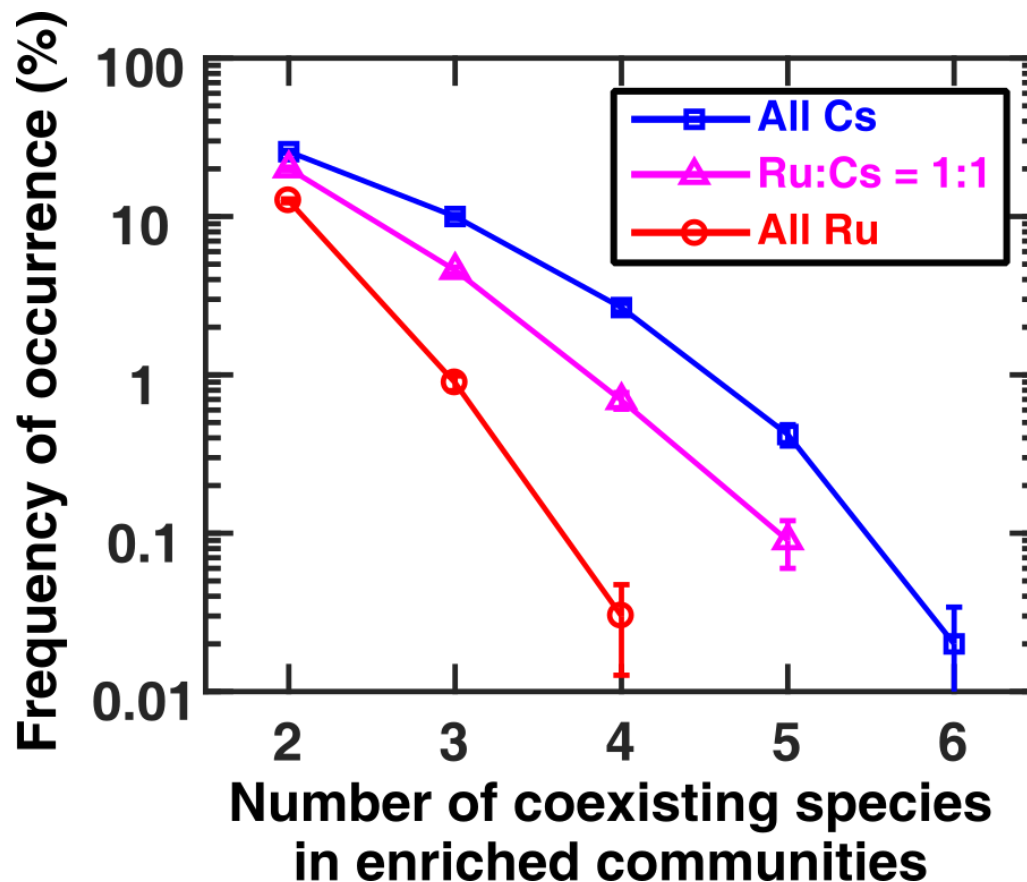
- Procedure:
  1. Simulate the enrichment process to get coexisting species (~200 gen.)



2. Repeat, using other parameters of the initial pool (randomly) to make an ensemble of communities that show species coexistence
3. Look for commonalities in network properties of coexisting species

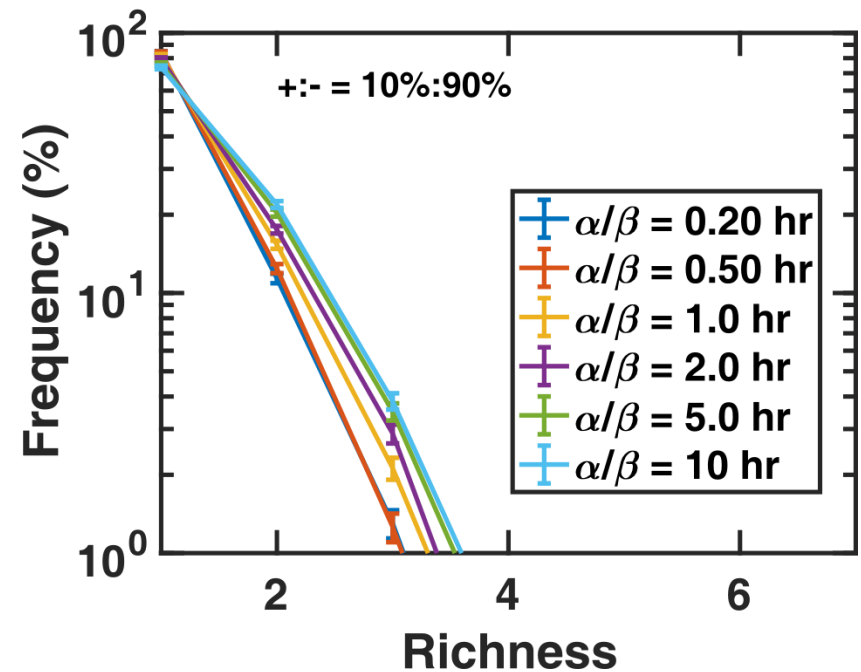
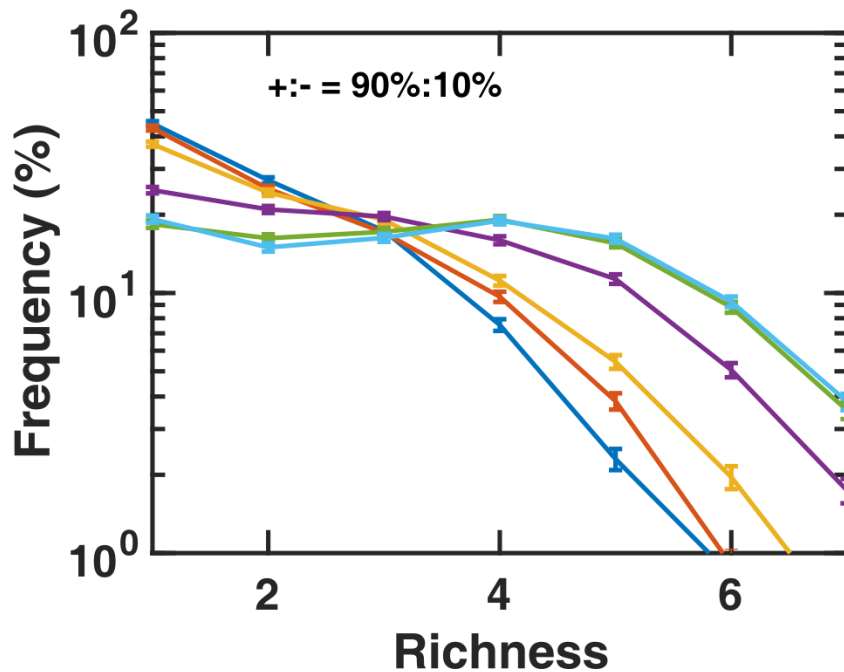
# Interaction mechanisms affect coexistence outcomes

- Comparing the same network of interactions with either consumable or reusable mediators



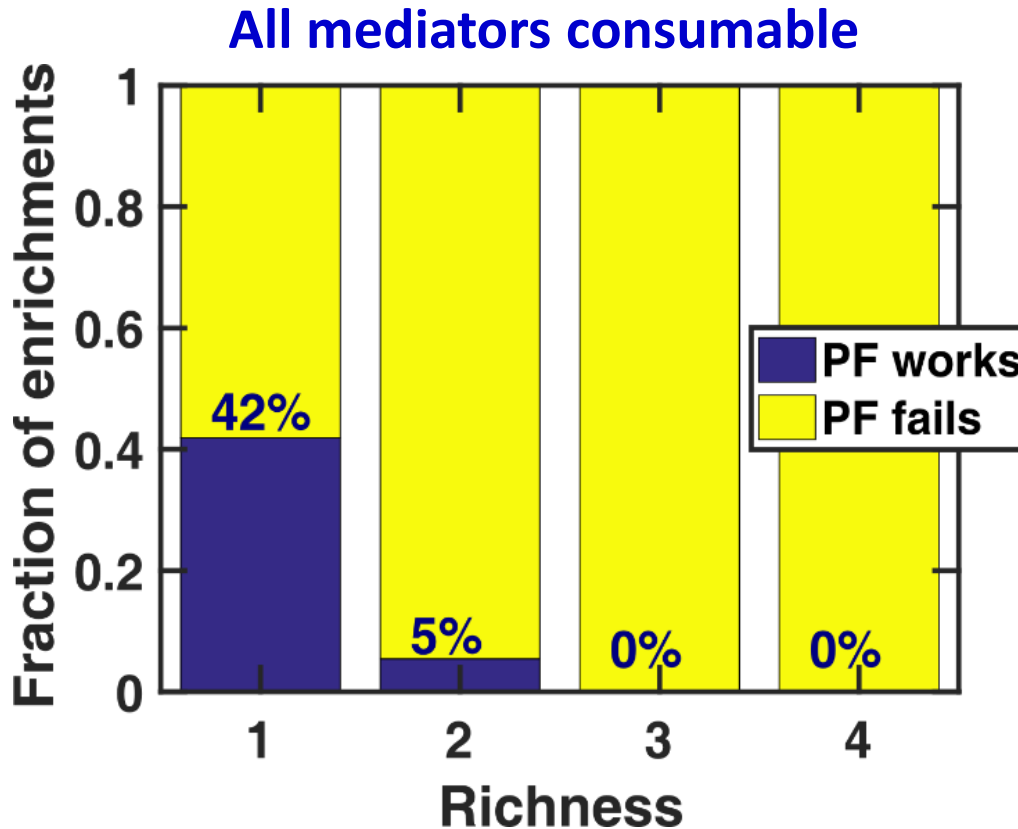
# Interaction mechanisms affect coexistence outcomes

- Consumption/degradation of mediators by cells has a pronounced impact on coexistence



# LV model fails to accurately predict coexistence

- Comparing coexistence predictions between a reference mediator-explicit model and a corresponding pairwise model



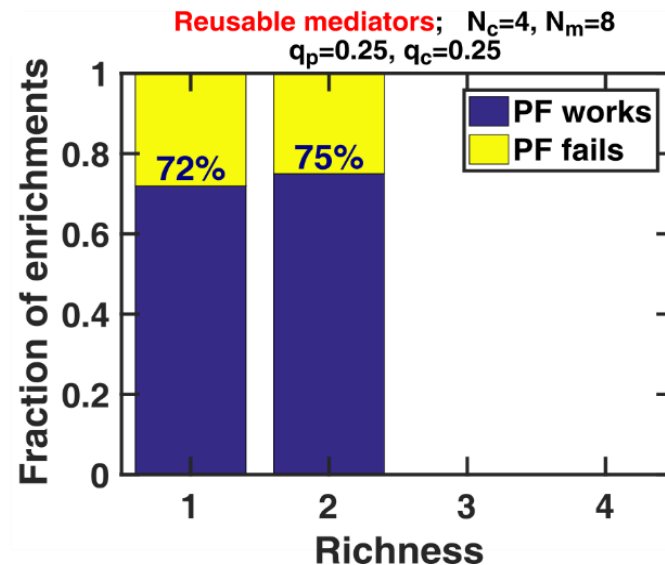
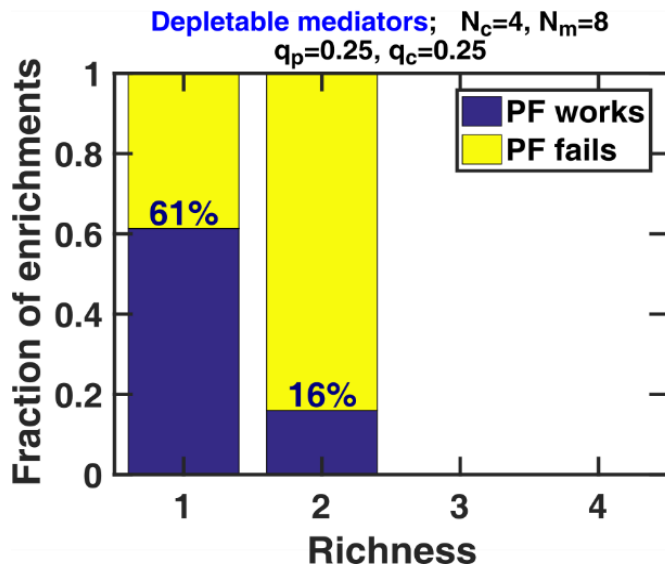
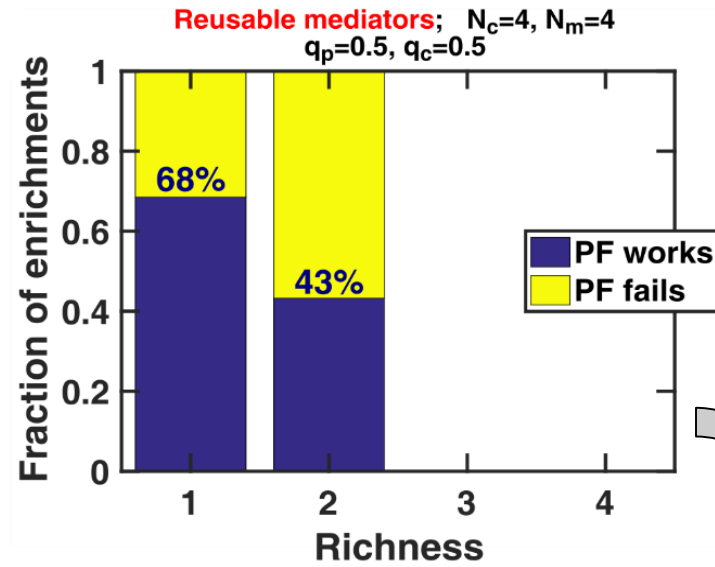
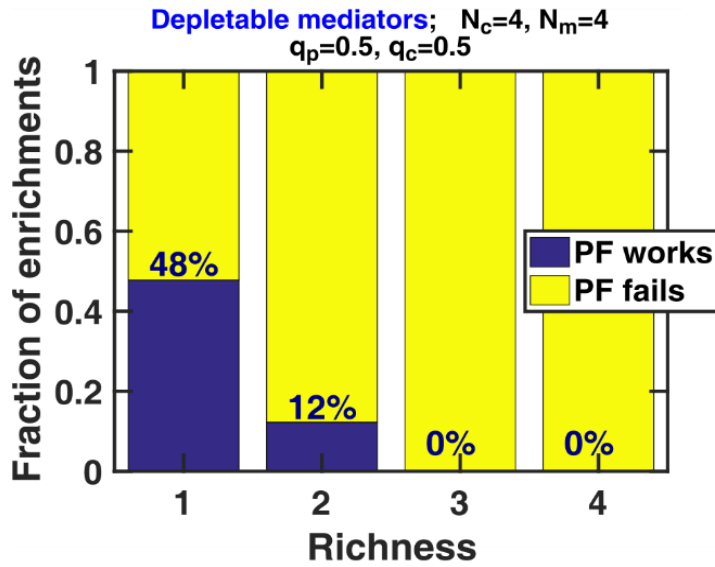
- Are there conditions under which LV works well?



# LV model predicts coexistence under certain conditions

What matters?

More reusable mediators



More independent interactions

# Summary

- A mediator-explicit model resolves some of the issues of pairwise Lotka-Volterra models, allowing a more realistic study of coexistence
- Interaction mechanisms (e.g. mediation through a reusable or a consumable chemical) seem to be important for coexistence predictions; thus motivating more mechanistic studies of interactions
- Pairwise LV models fail to predict coexistence when
  - Diverse interactions cannot be represented by a single LV equation
  - Interactions are not independent because of shared mediators
- What to do then?

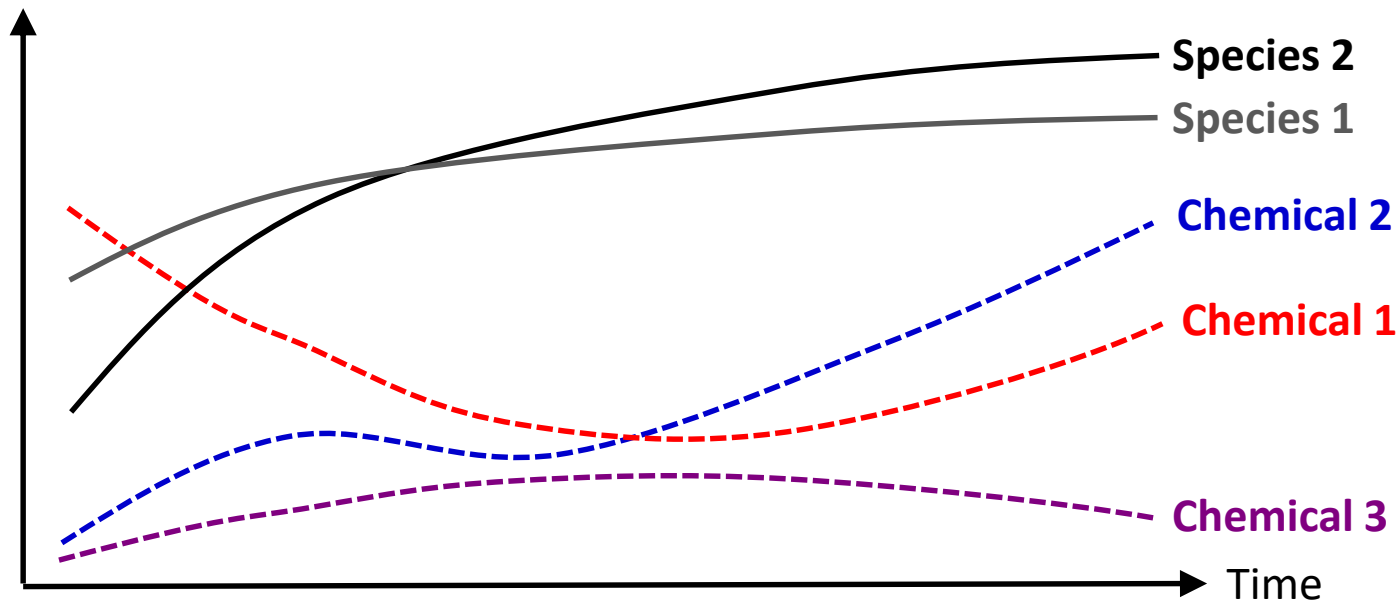
# Characterizing chemical-mediated interactions

Q1. Are there general equations that would adequately capture common chemical-mediated interactions?

Q2. How often are mediators shared (and interactions are interdependent)?

→ Systematically surveying chemical-mediated interactions

- Chemical profiling (colorimetric/fluorimetric assays, MS, NMR, etc.)
- **Challenge:** unclear *a priori* what chemicals are influential



# Coordinating the efforts to model microbial interactions

## 1. Database of microbe-microbe interactions

- Ongoing (to be made public soon)

## 2. Accessible user interface for simulating known mechanisms

- Engaging both theorists and experimentalists

## 3. Aggregation of raw interaction assay data (species + chemicals)

- What is the proper format?

# Thank you!

## Questions?



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